



MICROCOPY RESOLUTION, TEST CHART

SECURITY CLASSIFICATION OF THIS PAGE (When Date &

REPORT DOCUMENTATION PAGE	READ INSTRUCTIONS BEFORE COMPLETING FORM
T. REPORT NUMBER 1 AD-A14637	3. RECIPIENT'S CATALOG NUMBER
4. TITLE (and Aubitto) Enhancement of Immune Responses to Polysaccharide Antigens Using 8-Mercaptoguanosine	S. TYPE OF REPORT & PERIOD COVERED
7. AUTHOR(a) James J. Mond	8. CONTRACT OR GRANT NUMBER(2) NOO014-83-WBMC438
5. PERFORMING ORGANIZATION NAME AND ADDRESS Department of Medicine, USUHS 4301 Jones Bridge Road Bethesda, MD 20814	16. PROGRAM ELEMENT, PROJECT, TASK AREA & WORK UNIT NUMBERS NR 666-032
11. CONTROLLING OFFICE NAME AND ADDRESS Jeannine A. Majde, Ph.D. Scientific Officer, Immunology Code 441, Cellular Biosystems Group, Dept. of the Navy, ONR Arlington, VA 22217	12. REPORT DATE September 30, 1984 13. NUMBER OF PAGES 8
Same as #11.	15. SECURITY CLASS. (of this report) Unclassified 15. DECLASSIFICATION/DOWNGRADING SCHEDULE
16. DISTRIBUTION STATEMENT (of this Report) Unlimited DISTRIBUTION STATEMENT A	<u>L</u>

Approved for public relea n Unlimited

17. DISTRIBUTION STATEMENT (of the abetract entered in & Unlimited

10. SUPPLEMENTARY HOTES

19. KEY WORDS (Continue on reverse side if necessary and identify by block number) 8-mercaptoguanosine (8sGuo), TNP-Ficoll, pneumococcal polysaccharides, xid immune defect, ontogeny immune responsiveness, serum IgG, IgG, IgG, antibody.

STRACT (Continue on reverse side if necessary and identify by block number)

8-mercaptoguanosine (8sGuo) is an adjuvant that can be used to enhance the in vivo response of mice to polysaccharide antigens. This enhancement of the immune response is observed in all of the IgG subclasses examined, IgG, IgG, and IgG3. Most interestingly injection of this adjuvant restores the responses of nonresponder immune defective CBA/N mice to TNP-Ficoll the prototype

DD 1 JAN 72 1473

EDITION OF 1 NOV 65 IS DESCLETE

S/N 0102- LF- 014- 6601

SECURITY CLASSIFICATION OF THIS PASE (Then Date Bries

09 28 084

polysaccharide antigen we use in these experiments. Concomitant administration of this adjuvant together with TNP-Ficoll also enhanced the antibody responses of low responder neonatal mice to this antigen. Since our results suggested that this immune adjuvant was being rapidly metabolized in vivo we determined whether continuous injection of this adjuvant was more effective than a single dose. Our results indicate that injection of 8sGuo on three consecutive days was significantly more effective in enhancing anti-TNP-Ficoll responses than administration on one day only. We have determined from in vitro experiments that 8sGuo is exerting its predominant effect on activated and not resting B cells. Taken together our data suggest that this immune adjuvant will serve as a useful tool to investigate the regulatory controls of immune responses to polysaccharide antigens in normal and immune defective mice as well as in neonatal mice.

į	Accession For
	NTIS GRAMI
	D110 1110
	Unannounced [
1	Justification
	By
	Availability Codes
SOPY PECTED 9	Dist Special

Annual Report - 1983-1984

ONR Contract Number: NOOO14-83-WRM2408

- Enhancement of In Vivo Responses to Polysaccharide antigens with 8-mercaptoguanosine (8sGuo)
 - A. Responses to trinitrophenylated AECM-Ficoll. In view of the published reports by Goodman and Weigle demonstrating enhancement of antibody responses to TNP-Ficoll <u>in vitro</u> using 8sGuo we initiated a series of experiments to determine the effects of this adjuvant <u>in vivo</u>. When antigen and adjuvant are injected simultaneously there is on average a 10-100 fold increase in the serum IgG, anti-TNP titer seven days later (Table I).

Table I. 8sGuo enhances the in vivo antibody response to TNP-Ficoll.

	<u>IgG</u>	<u>IgG</u>	<u>IgG</u>	<u>IgM</u>
TNP-Ficoll +8sGuo	160 (1.38)	58 (1.49)	18 (1.37)	260 (1.19)
	975 (1.34)	831 (1.29)	142 (1.60)	283 (1.15)

The enhancement seen in the IgG_1 and IgG_3 antibody subclasses was not as dramatic as that seen in the IgG_2 subclass but was significant and reproducible. The peak antibody response was always noted seven to ten days after immunization and by day fourteen was declining (Table II).

Table II. Kinetics of the 8sGuo induced enhancement of the antibody response to TMP-Ficoll.

	Day 7	<u>Day 10</u>	<u>Day 14</u>	
		IgG_ anti-TNP titer		
TNP-Ficol1	180 (1.68)	124 (1.13)	117 (1.32)	
TNP-Ficoll + 8sGuo	1,108 (1.21)	517 (1.60)	379 (1.47)	

1. Groups of 6 mice were injected with TNP-Ficoll (10 μ g) and 8sGuo and sera obtained 7, 10 and 14 days later. Results represent geometric mean (γ / ÷log s.e.).

Whether the antigen was administered by intravenous or intraperitoneal route appeared to have little effect on the ability of 8sGuo to effect an enhanced response, however the enhancement was very dependent on the amount of 8sGuo injected (Table III).

Table III. 8sGuo induced enhancement of the IgG anti-TNP antibody response to TNP-Ficoll requires a high dose of adjuvant.

	Dose of 8sGuo	Day 7	<u>Day 10</u>
		IgG. anti-	INP-titer
TNP-Ficoll	-	42 (1.38)	368 (1.23)
TNP-Ficoll	30 mg	444 (1.27)	409 (1.23)
	10 mg	81 (1.39)	58 (1.23)
	3 mg	68 (1.19)	-

1. Group of 6 CBA/J mice were injected with TNP-Ficoll (10 μg) together with 30 mg, 10 mg or 3 mg of 8sGuo. Sera was obtained 7 and 10 days later.

Thus injection of less than 30 mg of 8sGuo per mouse was only marginally effective in enhancing the anti-TNP-Ficoll response, while doses of 50 - 100 mg per mouse were no more effective than 30 mg. This observation suggested to us the possibility that the adjuvant may be rapidly consumed in vivo and therefore large doses and possibly continuous administration of the adjuvant may be the most effective way to elicit an enhanced response. To test this we injected TNP-Ficoll on day 0 and 8sGuo on five consecutive days thereafter and compared its effect to that seen when the adjuvant is given on one day only (Table IV).

Table IV. Repeated injection of 8sGuo is more effective than a single injection in enhancing the anti-TNP-Ficoll response.

	IgG, Anti-T	NP-Titer IgG
TNP-Ficol1	863 (1.26)	141 (1.08)
TNP-Ficoll + 8sGuo (Day 0)	5,163 (1.05)	2,670 (1.14)
TNP-Ficoll + 8sGuo (Dav 0 - Dav 5)	19,229 (1.11)	27.521 (1.13)

l. Groups of 6 mice were injected with $10\mu g$ of TNP-Ficoll only, or together with 8sGuo or were given 8sGuo on days 0 through day 5. Sera were obtained seven days later and titered for the presence of anti-TNP-antibody.

From other data it appears that administration of the adjuvant on day 0 - day 3 was also significantly more effective in enhancing the response to TNP-Ficoll than when it was given on one day only.

B. Responses to pneumococcal polysaccharide type 14 (P-14). Having established that responses to a haptenated-polysucrose molecule (i.e.; TNP-Ficoll) could be readily enhanced by 8sGuo, we attempted to test another purified polysaccharide antigen (P-14) (Table V).

Table V. Enhancement of the IgG, and IgG, antibody responses to pneumococcal polysaccharide type 14 by 8sGuo.

	<u>IgG</u>	<u>IqG</u>
	anti-P-	·14-titer
P-14	<10	24 (1.24)
P-14 + 8MG	160 (1.40)	296 (1.30)

1. Groups of 6 mice were injected with 20 μg of pneumococcal polysaccharide type 14 (P-14) and sera obtained seven days later.

Responses to this antigen in the absence of adjuvant were very low and often below our level of detectability. In the presence of 8sGuo, however, this antigen stimulated a significant response which was always greater than that seen in the absence of adjuvant. Although the maximum

enhancement was seen when antigen and adjuvant were given together, significant enhancement of the response was observed even when antigen was injected seven days prior to the adjuvant (Table VI).

Table VI. 8sGuo enhances IgG, anti-P-14 response even when injected seven days after injection of the antigen.

					IgG.	anti-	-P-14 titer
P14	(day	- 7)				39	(1.46)
P14	(day	- 7)	+8sGuo	(day	0)	148	(1.40)
P14	(day	0)		_		37	(1.50)
P14	(day	0)	+8sGuo	(day	0)	622	(1.45)

1. Groups of 6 mice were injected with 20 μ g of Pl4 either 7 days prior to injection of 8sGuo (day - 7) or simultaneously with 8sGuo (day 0). Sera was obtained seven days after injection of 8sGuo.

Similar degrees of enhancement were observed when a mixture of pneumococcal polysaccharides (Pneumovax) were injected (data not shown).

II. Elicitation of in vivo responses to TNP-Ficoll in xid immune defective CBA/N mice.

PARTICLE CANADAN MUNICIPAL CANADANA SOCIALISMO CONTRACTOR CONTRACTOR

CBA/N mice carry an x-linked immune defect which precludes them from responding to polysaccharide antigens. In this regard they resemble the immune system of newborn mice which are also hyporesponsive to this group of antigens. To test whether 8sGuo could convert a "non-responder" strain into a "responder" strain, we immunized CBA/N mice with TNP-Ficoll and 8sGuo. We observed that administration of 8sGuo on five consecutive days to these mice restored the anti-TNP-Ficoll responses to a level which was comparable to that seen in control responder mice (Table VII).

Table VII. CBA/N mice with the \underline{xid} immune defect respond to TNP-Ficoll in the presence of 8sGuo.

	Anti-TNP-Titer				
	IgG ₁		<u>Iq</u> G	,	
	<u>Day 10</u>	Day 14	Day 10	Day 14	
TNP-Ficoll	<5	<5	<5	<5	
TNP-Ficoll +8sGuo	361 (1.07)	680 (1.15)	1,272 (1,10)	467 (1.13)	

1. Groups of 6 CBA/N x DBA/2 male mice were injected with TNP-Ficoll (10 μ g) on day 0 and with 8sGuo on day 0 through day 5. Results represent geometric mean (x/÷log s.e.).

This enhancement of responsiveness was seen in the ${\rm IgG}_1$ and ${\rm IgG}_2$ subclasses but not in the ${\rm IgM}$ and ${\rm IgG}_3$ subclasses of antibody.

III. In vitro enhancement of polysaccharide responses by 8sGuo

parameter personal parameter

A. Lack of requirement for continuous antigen. We wished to determine whether the 8sGuo induced enhancement of in vivo responses to TNP-Ficoll (Table VIII) was dependent on the continued presence of the antigen.

Table VIII. Enhancement of anti-TNP PFC response does not require the continuous presence of antigen.

"Priming" Antigen	24hrs	Medium 48hrs	72hrs	24hrs	8sGuo (0.25mM) 48hrs	72hrs
Medium	5(1.22)	12(1.95)	12(1.18)	30(1.15)	120(1.30)	130(1.01)
TNP-Ficoll 2.5ng/ml	17(1.54)	13(1.08)	9(2.34)	293(1.12)	598(1.07)	320(1.18)

"Primed" B Cells Cultured With

- 1. B cells from CBA/J were cultured for 48 hours at 5 x 10⁵ cells/ml with medium only or with TNP-Ficoll (2.5ng/ml) and then washed 3 times with Mishell Dutton medium and placed into culture with medium, 8-Mercaptoguanosine (0.25mM) or with 8-Mercaptoguanosine + TNP-Ficoll (2.5ng/ml) and then assayed for anti-TNP PFC 24, 48, or 72 hours later.
- Results represent geometric mean (x/÷ log s.e.) of triplicate wells.

was dependent on the continued presence of the antigen. To this end purified populations of murine B cells were cultured with TNP-Ficoll for 48 hours and then extensively washed. 8sGuo was added for an additional 48 hours and numbers of cells secreting anti-TNP antibody were enumerated. The number of anti-TNP PFC counted was equivalent to that seen when antigen and 8sGuo were cultured for the duration of the four day culture period. This suggested that the 8sGuo induced enhancement of the response to TNP-Ficoll did not require the continued presence of high concentrations of this antigen.

B. Rapid onset of action of 8sGuo. Since it was clear from our results as well as from those published reports of Goodman and Weigle that 8sGuo did not have to be added at the onset of culture, we wished to determine the minimum time that 8sGuo needed to be cultured with cells to exert its maximum effect (Fig. 1). B cells were cultured with antigen for 48 hours and 8sGuo added 12,24 or 48 hours prior to enumeration of anti-TNP plaque forming cells. Although enhancement of in vitro responses were observed even 12 hours after the addition of 8sGuo, the maximum enhancement was observed when 8sGuo was added for 48 hours prior to enumeration of antibody secreting cells.

CONTROL AND STATES AND SECOND REPRESENTANT

になったかから、大くなかがから

Although 8sGuo had been shown by Drs. Goodman and Weigle to be an effective polyclonal B cell activator it was not clear whether it was exerting its effect on all the B cells or only a subset of them. To address this issue we separated B cells using cell size as the criterion reflecting the degree of activation of the B cell. We have previously shown that elutriation of B cells was an effective technique that could be used to separate small (115 μ^3) B cells from large B cells (175 μ^3). Interestingly, 8sGuo

"resting" B cells. Furthermore while this adjuvant could induce a significant increase in the expression of B cell Ia, it was only marginally effective in stimulating such an increase in small resting B cells (data not shown). Thus it appears that 8sGuo is an effective B cell activator but only on B cells that have been previously activated either by intentional administration of antigen or by environmental antigen. Resting B cells in G appear to be resistant to the polyclonal B cell enhancing effects of 8sGuo.

D. <u>8sGuo is an effective adjuvant for neonatal B cells</u>. Having demonstrated that 8sGuo could enhance in vivo responses to TNP-Ficoll even in a non-responder strain of mice (CBA/N) we wished to determine whether responses of neonatal mice to this polysaccharide antigen were also enhanceable. Spleen cells from 3, 6 and 11 day old mice were cultured with TNP-Ficoll and 8sGuo (Table IX) and anti-TNP plaque forming cell responses enumerated 3 days later.

Table IX. 8-Mercaptoguanosine restores in vitro responsiveness of neonatal spleen cells to TNP-Ficoll.

Age of Mice	8-Mercaptoguanosine	TNP-Ficoll	TNP-Ficoll + 8-Mercaptoguanosine
3 days	30 (1.58)	3 (1.80)	114 (1.32)
6 days	76 (1.38)	0	222 (1.14)
ll days	73 (1.43)	3 (1.25)	190 (1.15)

Although B cells from these young mice are unresponsive to this antigen in the absence of 8sGuo they are responsive in its presence. Thus it appears that TNP-Ficoll could initiate the early steps of B cell activation even in immature B cells of neonatal mice and 8sGuo could exert its immuno-enhancing effects on these activated B cells.

Taken together these data provide strong evidence that the immune adjuvant 8sGuo will be useful as an approach to studying ways in which the immune response to even poor immunogens such as polysaccharides can be enhanced. Furthermore it gives one a tool to study how an immature immune system which is unresponsive to polysaccharides can be converted into a responder status.

DISTRIBUTION LIST

Immunological Defense Program

Annual, Final and Technical Reports (one copy each except as noted)

Fritz H. Bach, M.D. Director, Immunology Research Center University of Minnesota Box 724, Mayo Memorial Bldg. 420 Delaware St., SE Minneapolis, MN 55455

Francis A. Ennis, M.D.
Department of Medicine
University of Massachusetts
Medical School
55 Lake Avenue
Worcester, MA 01605

and the second of the second o

the country and the second of the second of

Fred D. Finkelman, M.D.
Department of Medicine
Uniformed Services University
of the Health Sciences
4301 Jones Bridge Road
Bethesda, MD 20814

Dr. Matthew J. Kluger
Department of Physiology
University of Michigan Med. School.
7620 Medical Science II Bldg.
Ann Arbor, MI 48109

Dr. Vijaya Manohar Burriston Laboratories, Inc. 5050 Beech Place Temple Hills, MD 20748

Dr. Ernest D. Marquez Bioassay Systems Corporation 225 Wildwor: Avenue Woburn, MA 01801

James J. Mond, M.D.
Department of Medicine
Uniformed Services University
of the Health Sciences
4301 Jones Bridge Road
Bethesda, MD 20814

Dr. Donna S. Sieckmann Infectious Diseases Program Center Naval Medical Research Inst. Mational Naval Medical Center Bethesda, MD 20814 Dr. John D. Clements
Department of Microbiology
and Immunology
Tulane University Medical Ctr.
1430 Tulane Avenue
New Orleans, LA 70112

Dr. Edward A. Havell Trudeau Institute P.O. Box 59 Saranac Lake, NY 12983

Dr. Arthur G. Johnson
Department of Medical
Microbiology and Immunology
University of Minnesota
School of Medicine
2205 East 5th Street
Duluth, MN 55812

Dr. Philip Lake Immunologic Oncology Division Lombardy Cancer Center Georgetown Univ. School of Med. Washington, DC 20007

W. John Martin, M.D., Ph.D. Laboratory, Dept. of Medicine Naval Hospital National Naval Medical Center Bethesda, MD 20814

Dr. Robert I. Mishell
Dept. of Microbiology &
Immunology
Univ. of California, Berkeley
Berkeley, CA 94720

Dr. Page S. Morahan Department of Microbiology Medical College of Pennsylvania 3300 Henry Avenue Philadelphia, PA 19129

Dr. Alan L. Schmaljohn
Department of Microbiology
University of Maryland
School of Medicine
660 W Redwood Street
Baltimore, MD 21201

Annual, Final and Technical Reports (Cont.)

David A. Stevens, M.D.

Department of Medicine

Santa Clara Valley Medical Center

Stanford University

751 S. Bascom Avenue

San Jose, CA 95128

Dr. Barnet M. Sultzer
Department of Microbiology &
Immunology
Downstate Medical Center
450 Clarkson Avenue
Brooklyn, NY 11203

Dr. Alvin L. Winters
Department of Microbiology
University of Alabama
University, AL 35486

Dr. Phyllis R. Strauss Department of Biology Northeastern University 360 Huntington Avenue Boston, MA 02115

G. Jeanette Thorbecke, M.D. Department of Pathology New York University School of Medicine 550 First Avenue New York, NY 10016

Lyn Yaffe, M.D.
Research Support Center
Naval Medical Research Inst.
National Naval Medical Center
Bethesda, MD 20814

Annual, Final and Technical Reports (one copy each except as noted)

Dr. Jeannine A. Majde, Code 441CB Scientific Officer, Immunology Program Office of Naval Research 800 N. Quincy Street Arlington, VA 22217

Administrator (2 copies)
Defense Technical Information Center
Building 5, Cameron Station
Alexandria, VA 22314

Annual and Final Reports Only (one copy each)

Commanding Officer Neval Medical Command Washington, DC 20372

Commanding Officer
Naval Medical Research & Development Command
National Naval Medical Center
Betheada, MD 20814

Director, Infectious Diseases Program Center Maval Medical Research Institute National Naval Medical Center Bethesda, MD 20814

Commander Chemical and Biological Sciences Division Army Research Office, P. O. Box 12211 Research Triangle Park, NC 27709

Commander
U.S. Army Research and Development Command
Attn: SGRD-PLA
Fort Detrick
Frederick, MD 21701

Commander USAMRID Fort Detrick Frederick, MD 21701

Directorate of Life Sciences Air Force Office of Scientific Research Bolling Air Force Base Washington, DC 20332

Administrative Contracting Officer
OWR Resident Representative
(address varies - obtain from Business Office)

Final and Technical Reports Only

Director, Naval Research Laboratory (6 copies) Attn: Technical Information Division, Code 2627 Washington, DC 20375

END

FILMED

11-84

DTIC